

### Three New Sesquiterpenoids from *Echinops ritro* L.

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From the whole plants of *E. ritro* L., the three new sesquiterpenoids ( $3\alpha,4\alpha,6\alpha$ )-3,13-dihydroxyguaia-7(11),10(14)-dieno-12,6-lactone (**1**), ( $3\alpha,4\alpha,6\alpha,11\beta$ )-3-hydroxyguaia-1(10)-eno-12,6-lactone (**2**), and (11 $\alpha$ )-11,13-dihydroarglanilic acid methyl ester (= ( $4\beta,6\alpha,11\alpha$ )-4,6-dihydroxy-1-oxoeudesm-2-en-12-oic acid methyl ester; **3**), together with eight known sesquiterpenoids, were isolated. Their structures were elucidated through analysis of spectroscopic data including extensive 2D-NMR.

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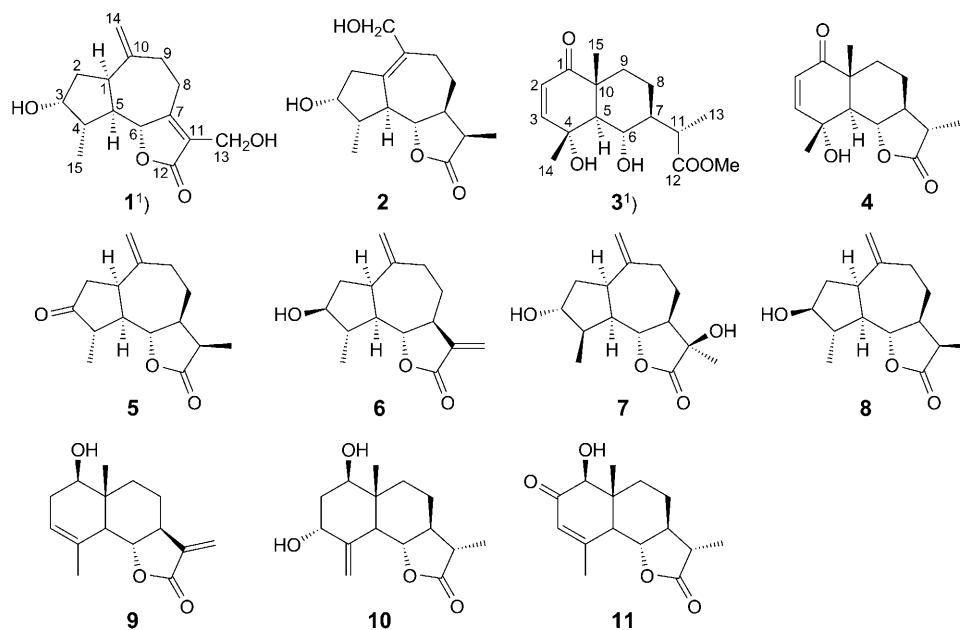
**Introduction.** – The genus *Echinops* (Compositae) consists of ca. 100 species in the world. This genus is represented by 17 species mainly distributed in northwestern part of China [1]. In China, *E. ritro* L. is only found in Xinjiang Uyghur Autonomous Region and is used as a remedy for different ailments in *Uighur Pharmacopeia* instead of *E. grijissi* HANCE (commercial name: Yuzhou Loulu) [2]. Previous chemical investigation on the *Echinops* species demonstrated the presence of polyacetylene-containing thiophenes [3–5], alkaloids [6], sesquiterpenoids [7], and flavone and flavone glycosides [8]. Further chemical analysis of *Echinops* species from Xinjiang Uyghur Autonomous Region led to the isolation of three new sesquiterpenoids, ( $3\alpha,4\alpha,6\alpha$ )-3,13-dihydroxyguaia-7(11),10(14)-dieno-12,6-lactone<sup>1)</sup> (**1**), ( $3\alpha,4\alpha,6\alpha,11\beta$ )-3-hydroxyguaia-1(10)-eno-12,6-lactone<sup>1)</sup> (**2**), and (11 $\alpha$ )-11,13-dihydroarglanilic acid methyl ester<sup>1)</sup> (**3**), along with the eight known compounds **4–11** [9–15] from the AcOEt extract of the MeOH extract. This article describes the isolation and structure elucidation of these new compounds.

**Results and Discussion.** – The IR spectrum of compound **1** indicated the presence of an OH group ( $3442\text{ cm}^{-1}$ ), an  $\alpha,\beta$ -unsaturated  $\gamma$ -lactone group ( $1737\text{ cm}^{-1}$ ), and a C=C bond ( $1658\text{ cm}^{-1}$ ). The molecular formula  $\text{C}_{15}\text{H}_{20}\text{O}_4$  was determined by HR-ESI-MS experiments ( $[M + \text{Na}]^+$  at  $m/z$  287.1249), in combination with  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR data (Table 1) indicating six degrees of unsaturation. Detailed analysis of the  $^1\text{H}$ ,  $^1\text{H}$ -COSY, HSQC, HMBC, and ROESY data established the structure of **1** as ( $3\alpha,4\alpha,6\alpha$ )-3,13-dihydroxyguaia-7(11),10(14)-dieno-12,6-lactone<sup>1)</sup>.

Analysis of the 1D-NMR data and HSQC spectra revealed that **1** contains one Me, three  $\text{CH}_2$ , and three CH groups, as well as an additional two CH groups and one  $\text{CH}_2$

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<sup>1)</sup> Trivial atom numbering; for systematic names, see *Exper. Part*.

Table 1.  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR Data (500 and 125 MHz, resp.;  $\text{CD}_3\text{OD}$ ) of **1**<sup>1</sup>) and **2**<sup>1</sup>).  $\delta$  in ppm,  $J$  in Hz.

	<b>1</b>		<b>2</b>	
	$\delta(\text{H})$	$\delta(\text{C})$	$\delta(\text{H})$	$\delta(\text{C})$
H–C(1)	3.22–3.27 ( <i>m</i> )	46.4		141.5
$\text{CH}_2$ (2)	1.89–1.92 ( <i>m</i> )	39.2	2.54–2.56 ( <i>m</i> ), 2.46–2.48 ( <i>m</i> )	40.5
H–C(3)	4.14–4.15 ( <i>m</i> )	75.4	4.04–4.06 ( <i>m</i> )	75.4
H–C(4)	2.25 ( <i>m</i> )	46.4	2.09–2.10 ( <i>m</i> )	45.7
H–C(5)	1.76–1.81 ( <i>m</i> )	50.8	2.47–2.49 ( <i>m</i> )	53.2
H–C(6)	4.69 ( <i>d</i> , $J = 11.5$ )	86.9	3.91 ( <i>t</i> , $J = 10.5$ )	86.7
C(7) or H–C(7)		170.4	2.33–2.38 ( <i>m</i> )	52.8
$\text{CH}_2$ (8)	3.29–3.30 ( <i>m</i> ), 2.35–2.45 ( <i>m</i> )	31.1	1.74–1.78 ( <i>m</i> ), 1.32–1.36 ( <i>m</i> )	26.2
$\text{CH}_2$ (9)	2.62–2.65 ( <i>m</i> ), 2.08–2.10 ( <i>m</i> )	33.9	2.45–2.46 ( <i>m</i> ), 2.10–2.12 ( <i>m</i> )	31.2
C(10)		150.4		136.5
C(11) or H–C(11)		126.1	2.64–2.67 ( <i>m</i> )	41.5
C(12)		175.0		182.1
$\text{CH}_2$ (13) or Me(13)	4.21–4.27 ( <i>dd</i> , $J = 18.5, 10.0$ )	53.7	1.11 ( <i>d</i> , $J = 7.5$ )	10.4
$\text{CH}_2$ (14)	5.19 ( <i>d</i> , $J = 3.5$ ), 4.90 ( <i>d</i> , $J = 3.5$ )	111.8	4.02	65.7
Me(15)	1.15 ( <i>d</i> , $J = 6.5$ )	16.0	1.19 ( <i>d</i> , $J = 6.5$ )	15.2

group attached to an O-function, an endocyclic C=C bond, an exocyclic  $\text{CH}_2=$  group, and one CO group (Table 1). The  $^1\text{H}$ -NMR spectrum showed the exocyclic  $\text{CH}_2=$  group at  $\delta(\text{H})$  5.19 and 4.90 (*d*,  $J = 3.5$  Hz, 1 H, each), an O-bearing  $\text{CH}_2$  group at  $\delta(\text{H})$  4.21–4.27 (*dd*,  $J = 18.5, 10.0$  Hz), and a Me group at  $\delta(\text{H})$  1.15 (*d*,  $J = 6.5$  Hz). These characteristic signals were in accordance with an guaian-type sesquiterpene skeleton. The  $^{13}\text{C}$ -NMR data of **1** were similar to those of ( $3\beta,4\alpha,11\beta$ )-3-hydroxyguai-10(14)-

eno-12,6-lactone [13], except for C(7), C(11), C(13), and C(3), which suggested they have similar structures. In the  $^{13}\text{C}$ -NMR spectrum, the signals of C(7) and C(11) were shifted downfield from  $\delta(\text{C})$  55.7 and 74.5 in [13] to  $\delta(\text{C})$  170.4 and 126.1 in **1**, which suggested that there is a C=C bond in **1** instead of single bond. The signal of C(13) was shifted downfield from  $\delta(\text{C})$  21.6 in [13] to  $\delta(\text{C})$  53.7 in **1**, meanwhile it was transformed into a CH<sub>2</sub> group from a Me group, which suggested that there is a CH<sub>2</sub>OH group in **1** instead of a Me group. This was further confirmed by the HMBCs CH<sub>2</sub>(2)/C(1), C(3), C(4), and C(5), H–C(4)/C(1), C(2), C(5), C(6), and C(15), H–C(5)/C(1), C(4), C(6), C(7), and C(15), CH<sub>2</sub>(14)/C(1) and C(9), CH<sub>3</sub>(15)/C(3), C(4) and C(5), and CH<sub>2</sub>(13)/C(7), C(11), and C(12), and the  $^1\text{H}, ^1\text{H}$ -COSY data confirmed the connectivity of C(1) to C(2) and C(5), of C(3) to C(2) and C(4), of C(5) to C(4) and C(6), and of C(8) to C(7) and C(9) (Fig. 1, a). The relative configuration of **1** was assigned by analysis of  $^1\text{H}$ -NMR coupling constants (Table 1) and ROESY correlations. A *J* value of ca. 11.5 Hz for H–C(6) suggested a *trans* configuration of H–C(6), assumed to be  $\beta$ -oriented, and H–C(5). The ROESY plot of **1** showed correlations of H–C(5) to H–C(1) and Me(15), of H–C(6) to H $_{\beta}$ -C(2) and H–C(4), and of H–C(3) to H $_{\beta}$ -C(2) and H–C(4) (Fig. 1, b). These findings showed that the relative configuration of H–C(1), H–C(3), H–C(4), H–C(5), and H–C(6) in **1** was  $\alpha$ ,  $\beta$ ,  $\beta$ ,  $\alpha$ , and  $\beta$ , respectively.

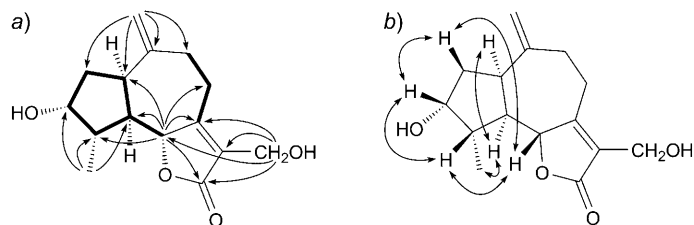


Fig. 1. a)  $^1\text{H}, ^1\text{H}$ -COSYs (—) and key HMBCs (H  $\rightarrow$  C) of **1**. b) Key ROESY correlations ( $\leftrightarrow$ ) of **1**.

A molecular formula C<sub>15</sub>H<sub>22</sub>O<sub>4</sub> was established for compound **2** from the HR-ESI-MS ( $[M + \text{Na}]^+$  at  $m/z$  289.1425). The  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR (Table 1) disclosed a guaiane-type skeleton [13]. The structure of **2** was established from the  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR,  $^1\text{H}, ^1\text{H}$ -COSY, HSQC, HMBC, and ROESY data as (3 $\alpha$ ,4 $\alpha$ ,6 $\alpha$ ,11 $\beta$ )-3-hydroxyguaia-1(10)-eno-12,6-lactone<sup>1</sup>.

The  $^{13}\text{C}$ -NMR spectrum of **2** showed chemical-shift values similar to those of (3 $\beta$ ,4 $\alpha$ ,11 $\beta$ )-3-hydroxyguaia-10(14)-eno-12,6-lactone [13], except for C(1), C(10), and C(14). The chemical shifts of C(1) ( $\delta(\text{C})$  141.5) and C(10) ( $\delta(\text{C})$  136.5) indicated the presence of a C=C bond between C(1) and C(10) in **2**. The signal of C(14) was shifted upfield from  $\delta(\text{C})$  111.2 in [13] to  $\delta(\text{C})$  65.7 in **2**, which suggested that a CH<sub>2</sub>OH group was present in **2** instead of a terminal CH<sub>2</sub>=C moiety. The structure of **2** was further confirmed by the HMBCs H–C(2)/C(1) and C(10), H–C(4)/C(2), H–C(5)/C(1) and C(10), H–C(6)/C(1), CH<sub>3</sub>(13)/C(7), and CH<sub>2</sub>(14)/C(1), C(9), and C(10), and by the  $^1\text{H}, ^1\text{H}$ -COSYs H–C(2)/H–C(3), H–C(5)/H–C(4), and H–C(8)/H–C(9) (Fig. 2, a). Finally, the relative configuration of **2** was established by a ROESY experiment and H-atom coupling constants. The H–C(6) (*t*, *J* = 10.5 Hz) suggested a *trans* configuration of H–C(6) with respect to H–C(5) and H–C(7). When H–C(6) was assigned as  $\beta$ -

oriented, the ROESY correlations (Fig. 2, b) of H–C(6) to H–C(4), H<sub>β</sub>–C(2), and Me(13) of H–C(5) to H–C(7) and Me(15), and of H–C(3) to H–C(4) and H<sub>β</sub>–C(2) suggested the depicted relative configuration of **2**.

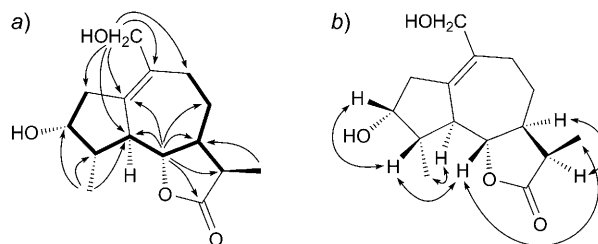


Fig. 2. a)  $^1\text{H}, ^1\text{H}$ -COSYs (—) and key HMBCs (H→C) of **2**. b) Key ROESY correlations (---) of **2**.

The HR-ESI-MS of compound **3** gave a quasi-molecular-ion peak at  $m/z$  297.1709 ( $[M + \text{H}]^+$ ), corresponding to the molecular formula  $\text{C}_{16}\text{H}_{24}\text{O}_5$ , requiring six degrees of unsaturation. The functional groups were also deduced from IR absorptions at 3312 (MeO), 1732 ( $\alpha,\beta$ -unsaturated 6-ring ketone group), and 1658 (C=C bond)  $\text{cm}^{-1}$ . Finally, the structure of **3** was established by its  $^1\text{H}, ^1\text{H}$ -COSY, HSQC, HMBC, and ROESY data as (11 $\alpha$ )-11,13-dihydroarglanilic acid methyl ester<sup>1</sup>).

The compound **3** probably originated from **4** through methanolysis during the plant extraction with MeOH. The  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR of data (Table 2) were very close to that of vulgarin (**4**) [9]. It showed the presence of the same eudesmane sesquiterpenoid skeleton, except for the presence of an MeO group ( $\delta(\text{C})$  51.9) in **3**. The functional groups were assigned on the basis of HMBC and  $^1\text{H}, ^1\text{H}$ -COSY studies (Fig. 3, a). The correlations of the MeO H-atoms to C(12) and C(11), and the correlations of H–C(6) to C(12) could not be found, which indicated that an acid ester was formed by opening

Table 2.  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR Data (500 and 125 MHz, resp.;  $\text{CD}_3\text{OD}$ ) of **3**<sup>1</sup>) and **4**<sup>1</sup>).  $\delta$  in ppm,  $J$  in Hz.

<b>3</b>			<b>4</b>	
	$\delta(\text{H})$	$\delta(\text{C})$	$\delta(\text{H})$	$\delta(\text{C})$
C(1)		205.5		201.6
H–C(2)	5.79 ( <i>d</i> , $J=10$ )	125.2	5.97 ( <i>d</i> , $J=10.5$ )	125.6
H–C(3)	6.61 ( <i>d</i> , $J=10$ )	155.2	6.68 ( <i>d</i> , $J=10.5$ )	151.7
C(4)		72.5		70.1
H–C(5)	2.03 ( <i>d</i> , $J=10.5$ )	56.6	2.54 ( <i>d</i> , $J=11.5$ )	55.0
H–C(6)	4.14 ( <i>t</i> , $J=10.5$ )	70.3	3.46 ( <i>t</i> , $J=11$ )	78.5
H–C(7)	1.62–1.63 ( <i>m</i> )	50.9	2.10–2.13 ( <i>m</i> )	48.2
CH <sub>2</sub> (8)	1.68–1.70 ( <i>m</i> ), 1.36–1.37 ( <i>m</i> )	22.3	1.89–1.92 ( <i>m</i> ), 1.60–1.64 ( <i>m</i> )	20.0
CH <sub>2</sub> (9)	1.88–1.90 ( <i>m</i> ), 1.36–1.37 ( <i>m</i> )	35.3	2.10–2.13 ( <i>m</i> ), 1.67–1.71 ( <i>m</i> )	34.2
C(10)		46.8		46.0
H–C(11)	2.94–2.97 (dq, $J=5.0, 6.0$ )	39.6	2.76–2.79 ( <i>m</i> )	37.8
C(12)		177.4		178.8
Me(13)	1.18 ( <i>d</i> , $J=6.0$ )	14.6	Me(13) 1.31 ( <i>d</i> , $J=8.0$ )	9.7
Me(14)	1.51 ( <i>s</i> )	23.2	1.80 ( <i>s</i> )	23.9
Me(15)	1.08 ( <i>s</i> )	20.0	1.28 ( <i>s</i> )	19.7
MeO	3.67 ( <i>s</i> )	51.9		

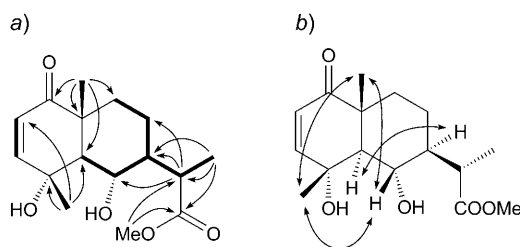


Fig. 3. a)  $^1\text{H},^1\text{H}$ -COSYs ( $\longrightarrow$ ) and key HMBCs (H  $\rightarrow$  C) of **3**. b) Key ROESY correlations ( $\longleftrightarrow$ ) of **3**

the lactone ring of **4**. ROESY Correlations were observed between H–C(5) and H–C(7), and between H–C(6) and Me(14) and Me(15) (Fig. 3, b), which confirmed that Me(14), Me(15), and H–C(6) were  $\beta$ -oriented. We determined the relative configuration of the Me group at C(11) as (11*S*) by comparison with the data of vulgarin.

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### Experimental Part

**General.** Column chromatography (CC) and TLC: silica gel ( $\text{SiO}_2$ ; 200–300 mesh) from Qingdao Marine Chemical Factory, Qingdao, P. R. China. Optical rotations: Jasco-DIP-370 digital polarimeter. M.p.: YuHua-X-4 apparatus. IR: Perkin-Elmer-577 spectrometer; KBr pellets; in  $\text{cm}^{-1}$ . 1D- and 2D-NMR Spectra: Bruker-AM-400 and -DRX-500 spectrometers;  $\text{Me}_4\text{Si}$  as an internal standard. MS: VG-Autospec-3000 mass spectrometer; in  $m/z$  (rel. %).

**Plant Material.** The whole plants of *E. ritro* L. were collected from Xinjiang Uyghur Autonomous Region, P. R. China, in August, 2006, and were identified by Prof. Sun Hang of the Kunming Institute of Botany, Chinese Academy of Sciences. A voucher specimen has been deposited with the Herbarium of the Kunming Institute of Botany, Chinese Academy of Sciences.

**Extraction and Isolation.** The air-dried and powdered sample (30 kg) was extracted exhaustively with MeOH (95%) under reflux and concentrated to give a crude extract which was suspended in  $\text{H}_2\text{O}$  and then extracted exhaustively with petroleum ether, AcOEt, and BuOH. The AcOEt fraction (160 g) was subjected to CC ( $\text{SiO}_2$  (200–300 mesh; 1.5 kg),  $\text{CHCl}_3$ , then 99  $\rightarrow$  90%  $\text{CHCl}_3/\text{MeOH}$ , then MeOH): Fractions 1–6. Compounds **1** (10 mg), **2** (19 mg), and **3** (35 mg) were obtained from Fr. 3 after repeated CC ( $\text{SiO}_2$ , petroleum ether/acetone), and RP-18).

(3*a*,4*a*,6*a*)-3,13-Dihydroxyguaia-7(11),10(14)-dieno-12,6-lactone (= rel-(6*a*R,8R,9*S*,9*a*R,9*b*R)-5,6,6*a*,7,8,9,9*a*,9*b*-Octahydro-8-hydroxy-3-(hydroxymethyl)-9-methyl-6-methyleneazuleno[4,5-*b*]furan-2(4H)-one; **1**): Colorless crystals from MeOH. M.p. 172–174°.  $[\alpha]_{\text{D}}^{25} = +20.9$  ( $c = 0.00575$ , MeOH). IR (KBr): 3442, 2933, 1737, 1658, 1641, 1016.  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR: Table 1. HR-ESI-MS (pos.): 287.1249 ( $\text{C}_{15}\text{H}_{20}\text{NaO}_4^+$ ; calc. 287.1259).

(3*a*,4*a*,6*a*,11*β*)-3-Hydroxyguaia-1(10)-eno-12,6-lactone (= rel-(3R,3*a*S,8R,9*S*,9*a*S,9*b*S)-3*a*,4,5,7,8,9,9*a*,9*b*-Octahydro-8-hydroxy-6-(hydroxymethyl)-3,9-dimethylazuleno[4,5-*b*]furan-2(3H)-one; **2**): Colorless crystals from MeOH. M.p. 167–169°.  $[\alpha]_{\text{D}}^{25} = +4.1$  ( $c = 0.0097$ , MeOH). IR (KBr): 3440, 2933, 1767, 1749, 1219, 984.  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR: Table 1. EI-MS: 266 (20,  $M^+$ ), 248 (100), 230 (30), 217 (35), 175 (60), 163 (90). HR-ESI-MS (pos.): 289.1425 ( $\text{C}_{15}\text{H}_{22}\text{NaO}_4^+$ ; calc. 289.1415).

(11*α*)-11,13-Dihydroarglanilic Acid Methyl Ester (= rel-( $\alpha$ R,1R,2R,4*a*S,8*S*,8*a*R)-1,2,3,4,4*a*,5,8,8*a*-Octahydro-1,8-dihydroxy- $\alpha$ ,4*a*,8-trimethyl-5-oxonaphthalene-2-acetic Acid Methyl Ester; **3**): Colorless

crystals from MeOH. M.p. 181–184°.  $[\alpha]_{\text{D}}^{21} = -42.7$  ( $c = 0.01475$ , MeOH). IR (KBr): 3312, 2983, 2950, 1732, 1678, 1252.  $^1\text{H}$ - and  $^{13}\text{C}$ -NMR: Table 2. EI-MS: 278 (4,  $[M-\text{H}_2\text{O}]^+$ ), 264 (7), 249 (80), 203 (45), 172 (100), 149 (60). HR-ESI-MS (pos.): 297.1709 ( $[M+\text{H}]^+$ ,  $\text{C}_{16}\text{H}_{25}\text{O}_5^+$ ; calc. 297.1701).

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